



**5<sup>th</sup> International Workshop on  
PET in Lymphoma  
Palais de l' Europe, Menton France  
September 19-20, 2014**

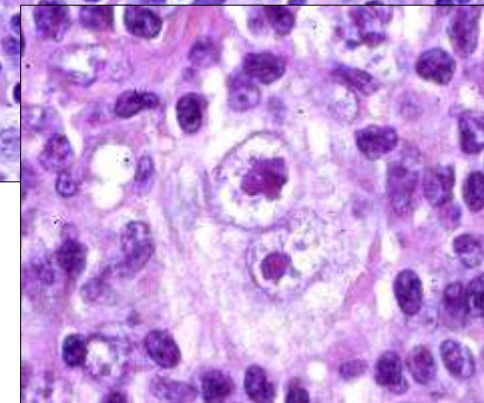
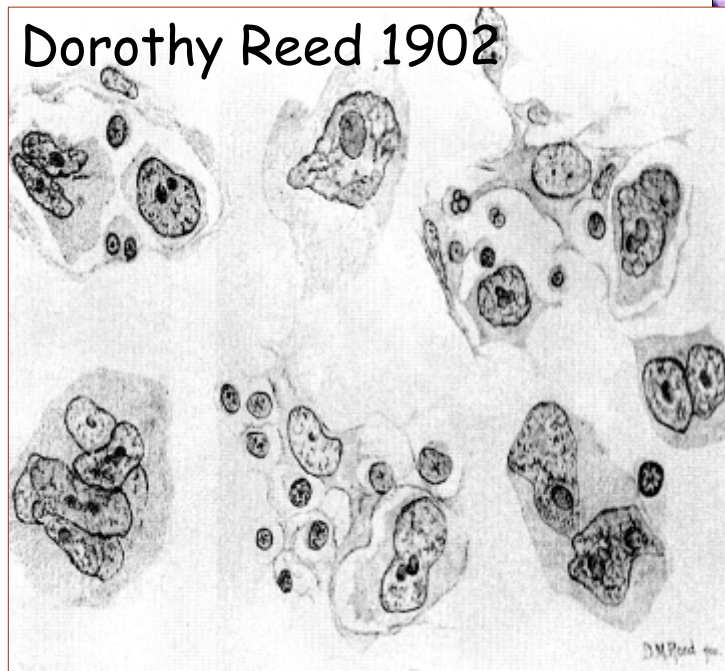
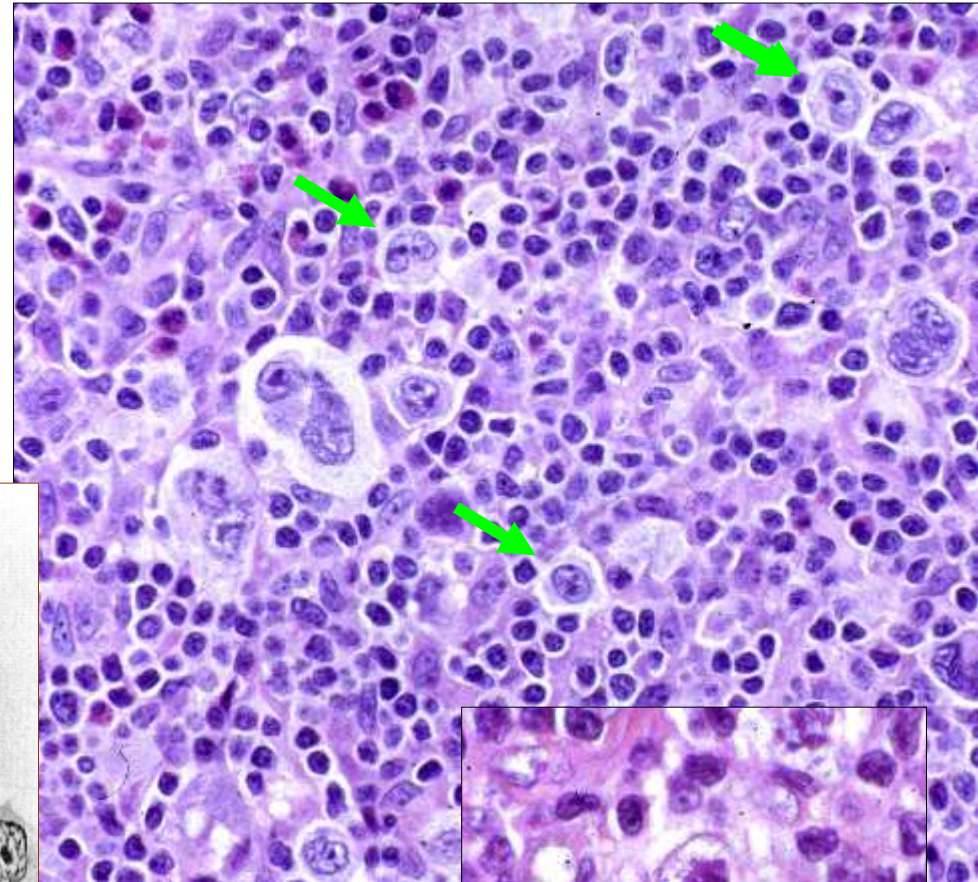
**A clinical-pathological algorithm based on  
the combination of interim PET with biological markers in  
Classical Hodgkin Lymphoma**



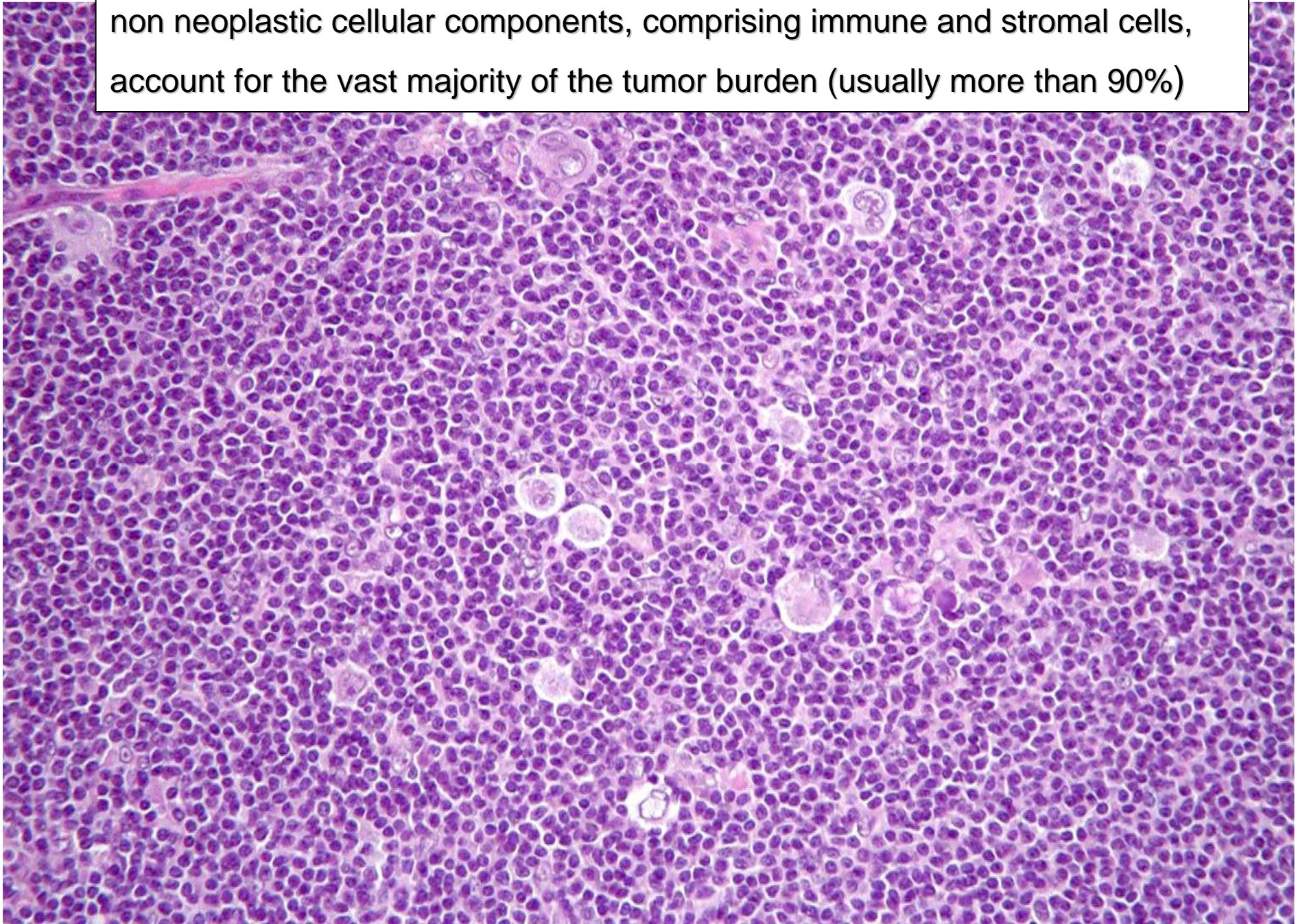
Anatomic Theatre – Bologna University

# Classical Hodgkin Lymphoma

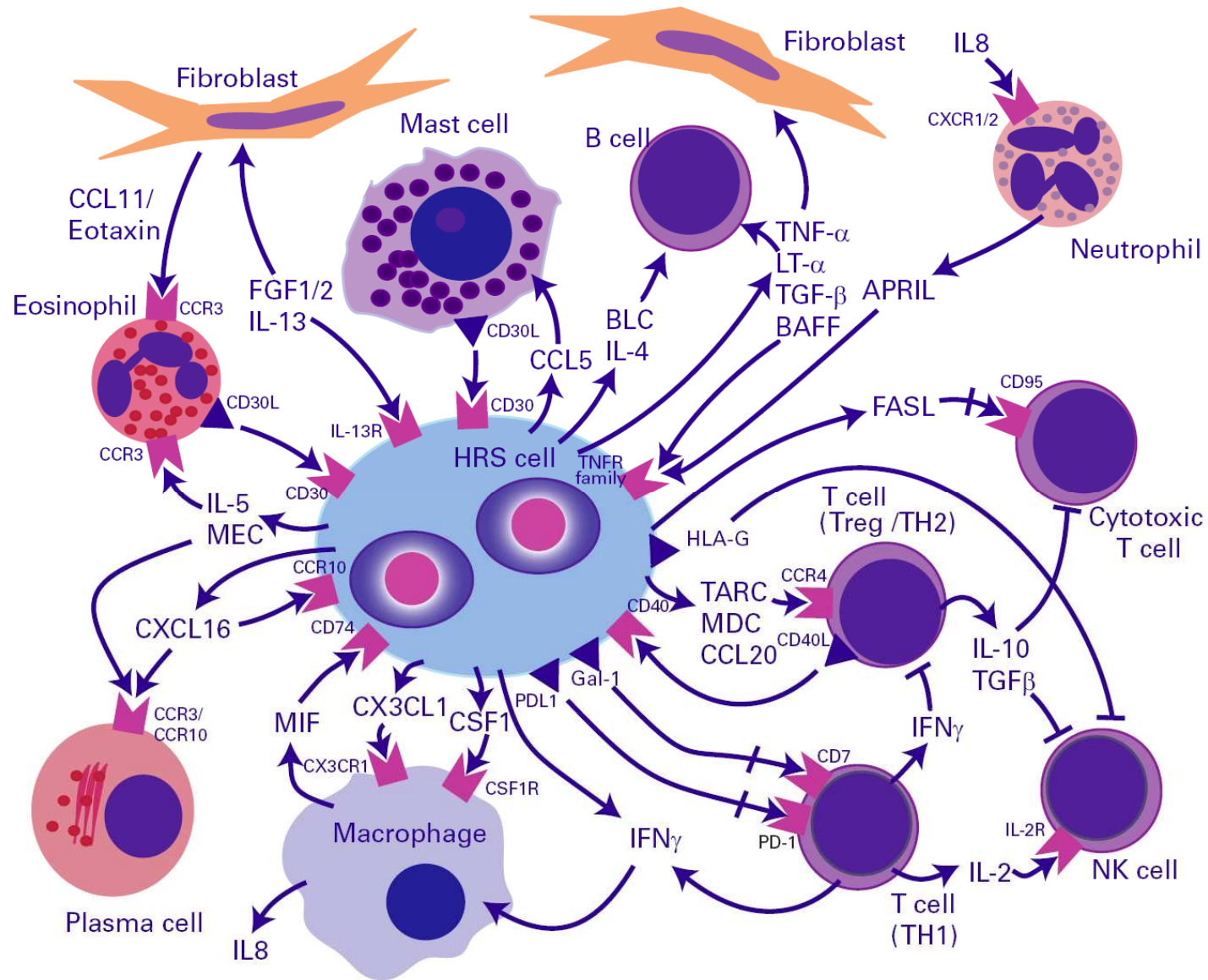
Classical Hodgkin Lymphoma is a monoclonal lymphoid neoplasm (in most instance derived from B cells) composed of mononuclear Hodgkin cells and multinucleated Reed-Sternberg cells



non neoplastic cellular components, comprising immune and stromal cells, account for the vast majority of the tumor burden (usually more than 90%)

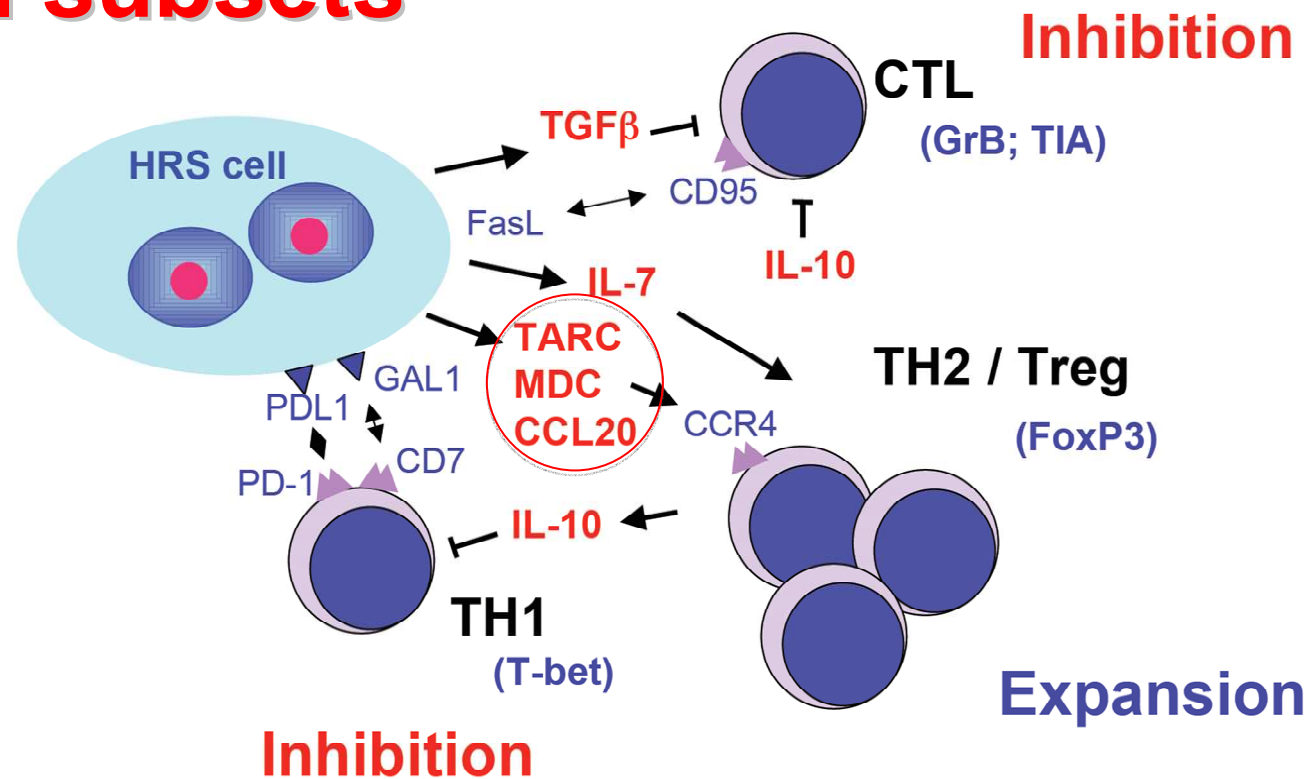


# CHL: cross-talk between HRSCs and microenvironment



Steidl C, Connors JM, Gascoyne RD. JCO 2011, 29:1812-26

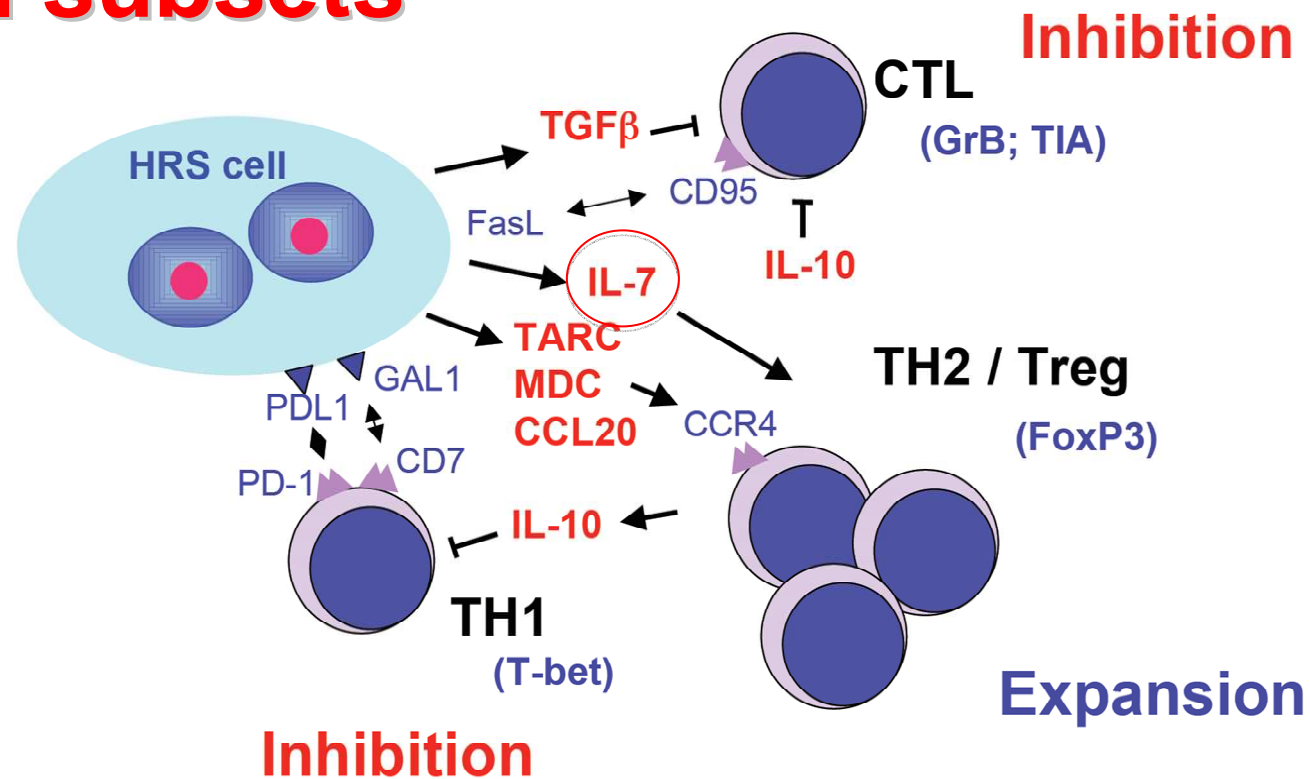
# T cell subsets



## HRS cells produce chemokines

- which attract TH2 and Treg cells
- capable of inducing differentiation of CD4 naive T cells toward FOXP3<sup>+</sup> T-reg cells
- which exert inhibitory effects on T-cell effector functions, especially on cytotoxic T lymphocytes

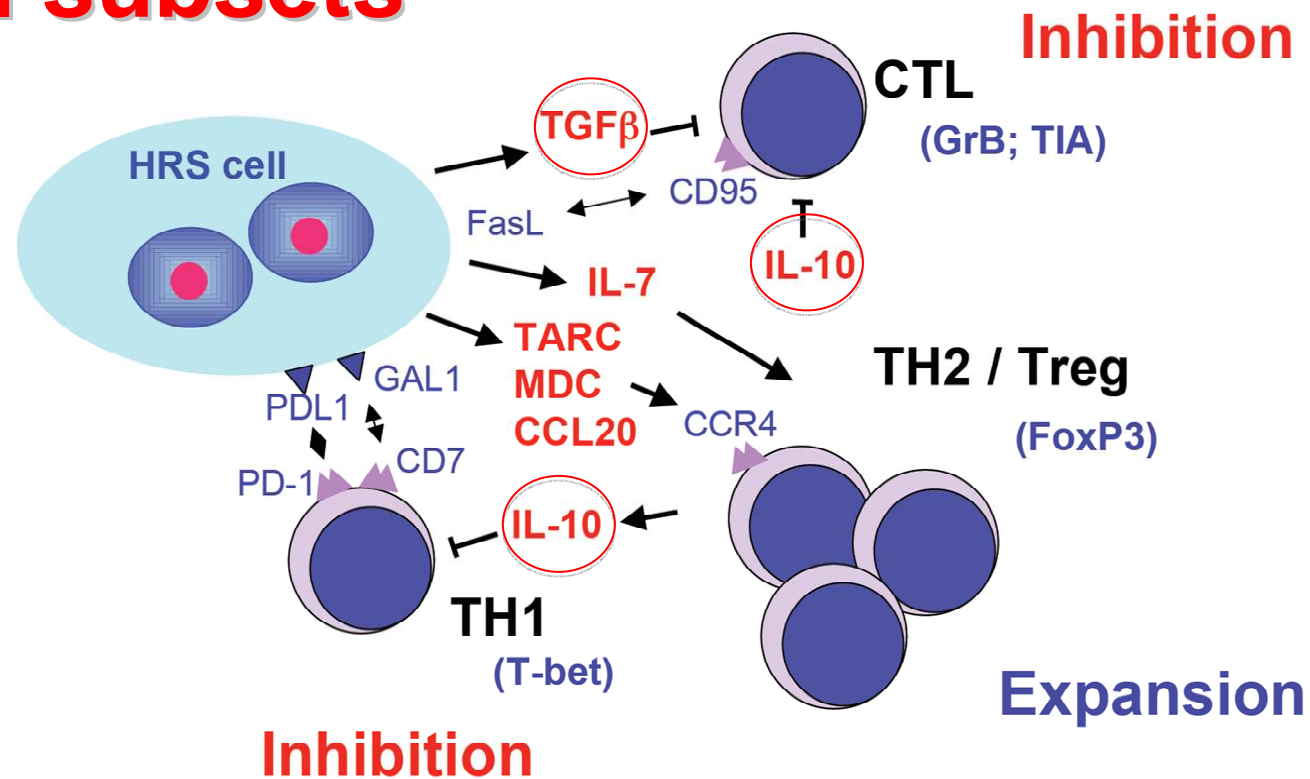
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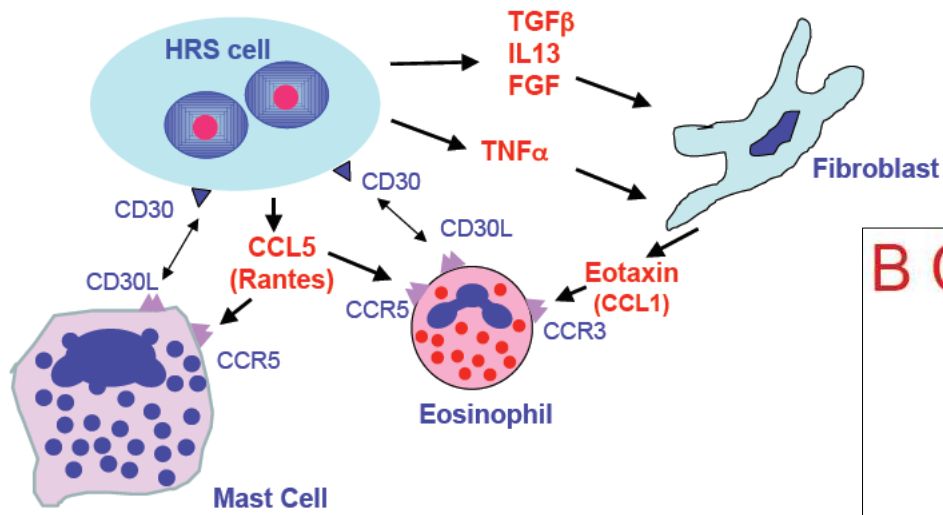
# T cell subsets



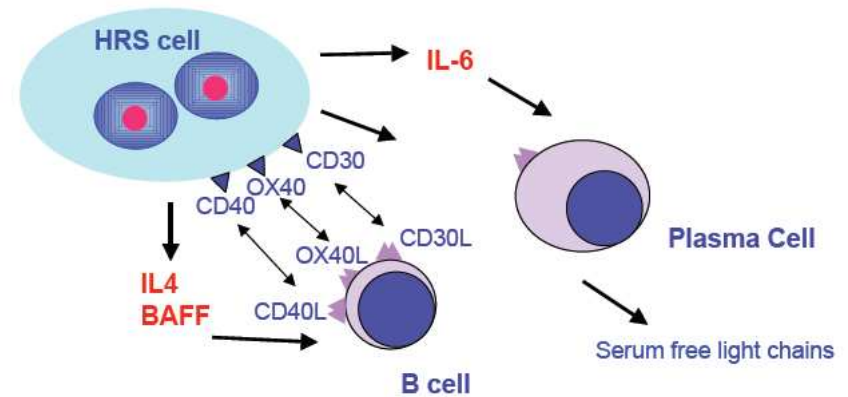
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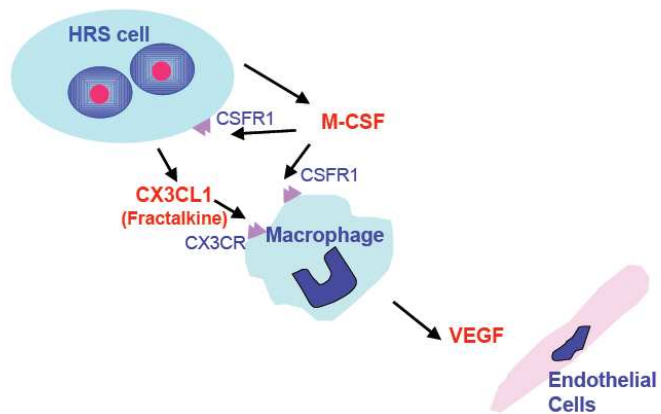
# Eosinophils and Mast Cells



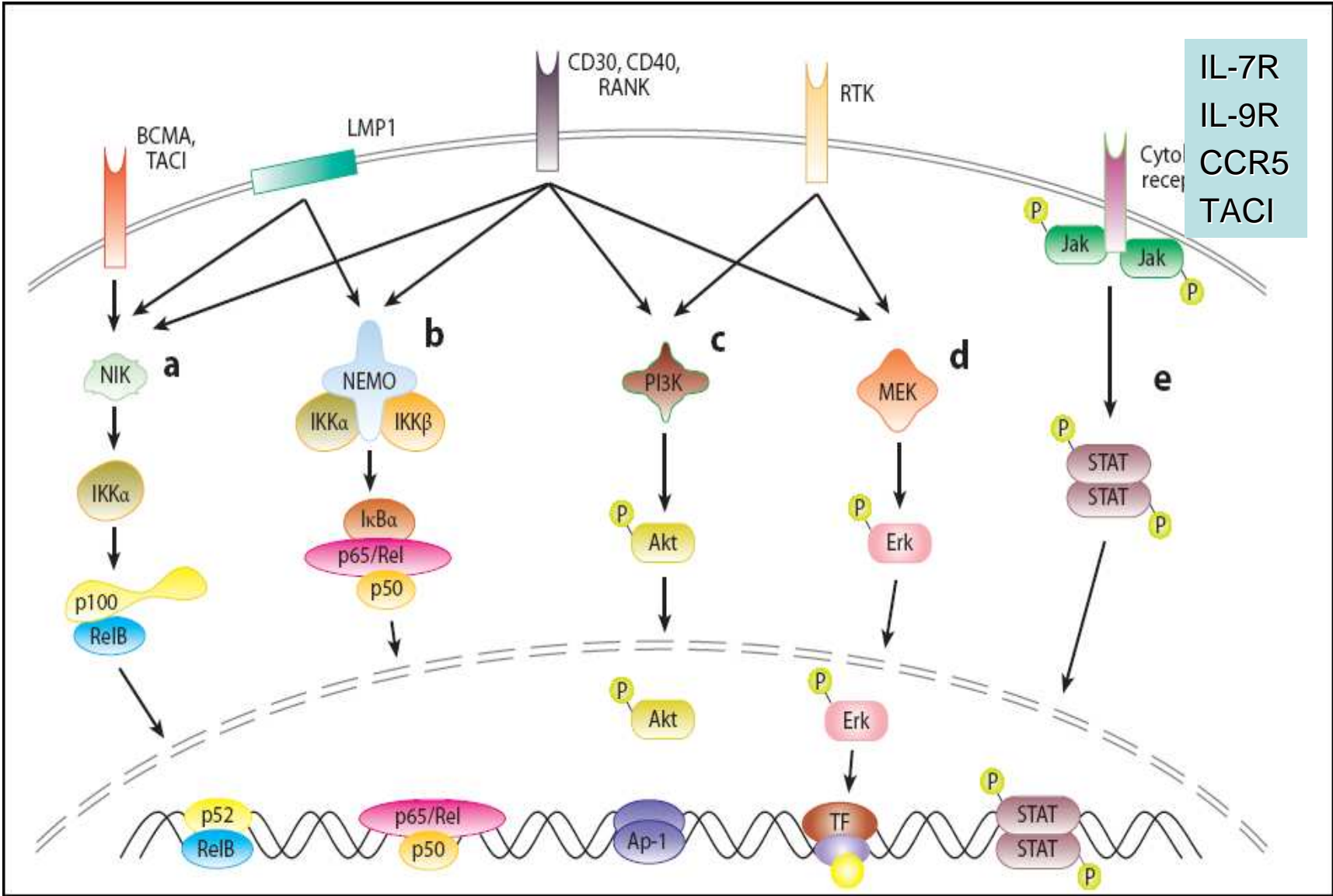
# B Cells and Plasma Cells



# Macrophages







Steidl C, Connors JM, Gascoyne RD. JCO 2011, 29:1812-26

| Immunohistochemistry                                |   |                              |  |               |
|---|---|------------------------------|--|---------------|
| Marker/Signature                                    | Expressed on/Staining Pattern   | Function                     | Outcome Correlation  | Reference No. |
| Granzyme B  | Cytotoxic T cells   | Target cell lysis            | Adverse (PFS, OS)  | 133-137       |
| TIA-1   | Cytotoxic T cells   | Target cell lysis            | Adverse (EFS, OS)  | 136,138       |
| FOXP3   | Regulatory T cells  | Transcriptional regulation   | Favorable (EFS)  | 137-139       |
| CD20  | Background B cells  | B-cell differentiation       | Favorable (OS, EFS)  | 132,140       |
| BCL11A  | Background B cells, plasmacytoid dendritic cells  | Transcriptional regulation   | Favorable (OS, EFS)  | 140           |
| HLA-DR  | HRS cells   | Antigen presentation         |  |               |
| CD68, PNA   | Macrophages   | Scavenger receptor           | Favorable (FFS)  | 123           |
| ALDH1A1   | Macrophages   | Oxidative pathway/metabolism | Adverse (PFS, DSS)   | 132,141       |
| STAT1   | Macrophages   | Transcriptional activation   | Adverse (DSS)  | 142           |
| EBV-encoded small RNAs                              | HRS cells   | Activation of NFκB           | Adverse in elderly patients, favorable in young patients (FFS, OS) | 143-147       |
| MMP11   | HRS cells, macrophages, endothelial cells, extracellular  | Tissue remodeling            | Adverse (PFS)  | 132           |
| Gene Expression Profiling                           |   |                              |  |               |
|   | Main Gene Components  |                              | Outcome Correlation  | Reference No. |
| Angiogenic signature                                | <i>ADH1B, CD93, SRPX, PLA2G2A, GPR126</i>   |                              | Adverse (primary treatment failure)                                | 132           |
| Adipocyte signature                                 | <i>GLUL, MGST1, COL1A2, FABP4</i>   |                              | Adverse (primary treatment failure)                                | 132           |
| Fibroblast function/extracellular matrix remodeling | Adverse: <i>MMP2, MMP3, TIMP1, COL1A1, COL4A1, COL4A2, COL5A1, COL18A1, COL16A1, MFAP2, THBS1/2, FN1, EDNRA, ITGB5, LAMA4</i> ; favorable: <i>TIMP4, SPON1, LAMB1, TACR1, CCL26</i> |                              | Discordant: adverse/favorable (primary treatment outcome)          | 142,148       |
| B-cell signature                                    | <i>BCL11A, BANK1, STAP1, BLNK, FCER2, CD24, CCL21</i>   |                              | Favorable (primary treatment outcome)                              | 132,140       |
| Cytotoxic T-cell signature                          | <i>CD3D, CD8B1, CTSL, CD26, SH2D1A, IFI16, RGS13, CR2, ELL3, CCDC23, PPM1L, TRA@, PIK3CA</i>  |                              | Adverse (primary treatment outcome)                                | 131,132,142   |
| Plasmacytoid dendritic cells                        | <i>ITM2A, SRPX, CTSB, APP</i>   |                              | Adverse (primary treatment outcome)                                | 132           |
| Macrophage signature                                | <i>ALDH1A1, LYZ, STAT1, ITGA4, CCL13, MS4A4A, CCL23, VCAN, HSP90AB3P, HSP90AB1, CTSB, CFL1, JMJD6, MAPK7, IKBKG, RAB7A, RXRA, MAPK13</i>  |                              | Adverse (primary treatment outcome)                                | 131,132,142   |

## Macrophages predict treatment outcome in Hodgkin's lymphoma

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**Haematologica 2010, 96:186-9**

| Markers used                          | Method         | #   | Outcome correlation  | Reference  |
|---------------------------------------|----------------|-----|--|--|
| PNA                                   | Histochemistry | 43  | Adverse (refractory disease, early relapse)                    | Ree <i>et al.</i> , Cancer 1985 <sup>10</sup>                                  |
| STAT1, ALDH1A1                        | GE, IHC        | 235 | Adverse (disease-specific survival)                            | Sanchez-Aguilera <i>et al.</i> , Blood 2006 <sup>11</sup>                      |
| LYZ, STAT1, ALDH1A1                   | GE, IHC        | 194 | Adverse (refractory disease, early relapse)                    | Sanchez-Espiridion <i>et al.</i> , Clinical Cancer Research 2009 <sup>13</sup> |
| CD68                                  | IHC            | 166 | Adverse (progression-free survival, disease-specific survival) | Steidl <i>et al.</i> , NEJM 2010 <sup>9</sup>                                  |
| LYZ, STAT1                            | GE             | 262 | Favorable (failure-free survival)                              | Sanchez-Espiridion <i>et al.</i> , Blood 2010 <sup>12</sup>                    |
| CD68, CD163                           | IHC            | 288 | Adverse (event-free survival, overall survival)                | Kamper <i>et al.</i> , Haematologica 2011 <sup>8</sup>                         |
| CD68                                  | IHC            | 59  | Adverse (refractory disease)                                   | Benedicte <i>et al.</i> , Blood 2010 [abstr.] <sup>34</sup>                    |
| CD68 (also in combination with FOXP3) | IHC            | 122 | Adverse (freedom from treatment failure, overall survival)     | Greaves <i>et al.</i> , Blood 2010 [abstr.] <sup>25</sup>                      |
| CD68                                  | IHC            | 144 | Adverse (event-free survival, disease-specific survival)       | Yoon <i>et al.</i> , Blood 2010 [abstr.] <sup>35</sup>                         |
| CD68                                  | IHC            | 105 | Adverse (overall survival)                                     | Tzankov <i>et al.</i> [personal communication]                                 |
| CD68                                  | IHC            | 45  | Adverse (progression-free survival)                            | Hohaus & Larocca [personal communication]                                      |
| CD68                                  | IHC            | 153 | Adverse (overall survival, progression-free survival)          | Farinha <i>et al.</i> [abstr.] <sup>36</sup>                                   |

PNA: peanut agglutinin, GE: gene expression (mRNA), IHC: immunohistochemistry.

# Interim PET (PET-2)

5%-12% PET2 negative pts experience a treatment failure

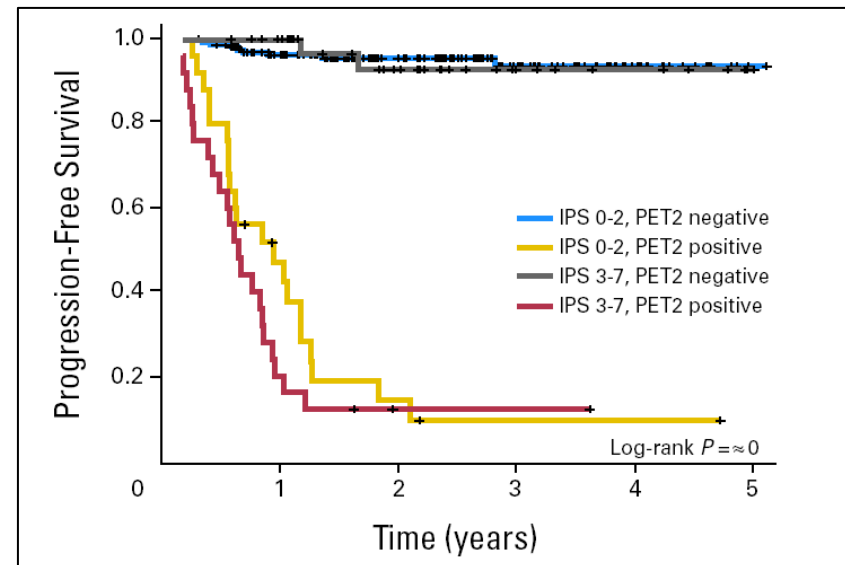
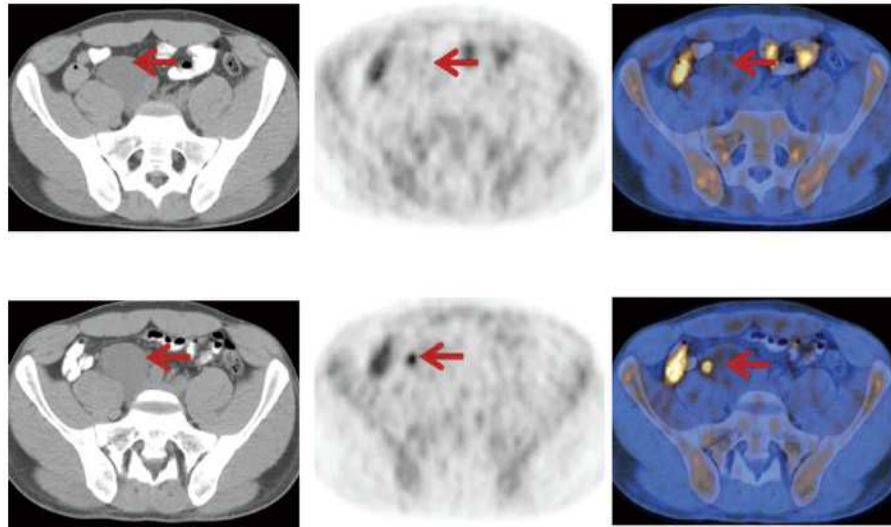
VOLUME 25 · NUMBER 24 · AUGUST 20 2007

JOURNAL OF CLINICAL ONCOLOGY

ORIGINAL REPORT

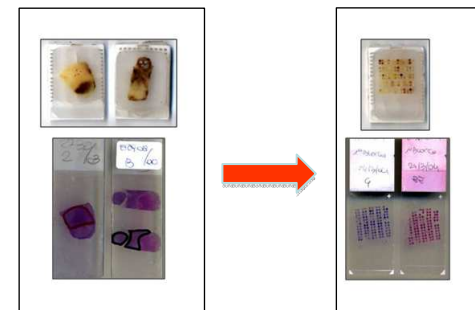
Early Interim 2-<sup>[18F]</sup>Fluoro-2-Deoxy-D-Glucose Positron Emission Tomography Is Prognostically Superior to International Prognostic Score in Advanced-Stage Hodgkin's Lymphoma: A Report From a Joint Italian-Danish Study

Andrea Gallamini, Martin Hutchings, Luigi Rigacci, Lena Specht, Francesco Merli, Mads Hansen, Caterina Patti, Amika Loft, Francesco Di Raimondo, Francesco D'Amore, Alberto Biggi, Umberto Vitolo, Caterina Stelitano, Rosario Sancetta, Livio Trentin, Stefano Luminari, Emilio Iannitto, Simonetta Viviani, Ivana Pierri, and Alessandro Levis

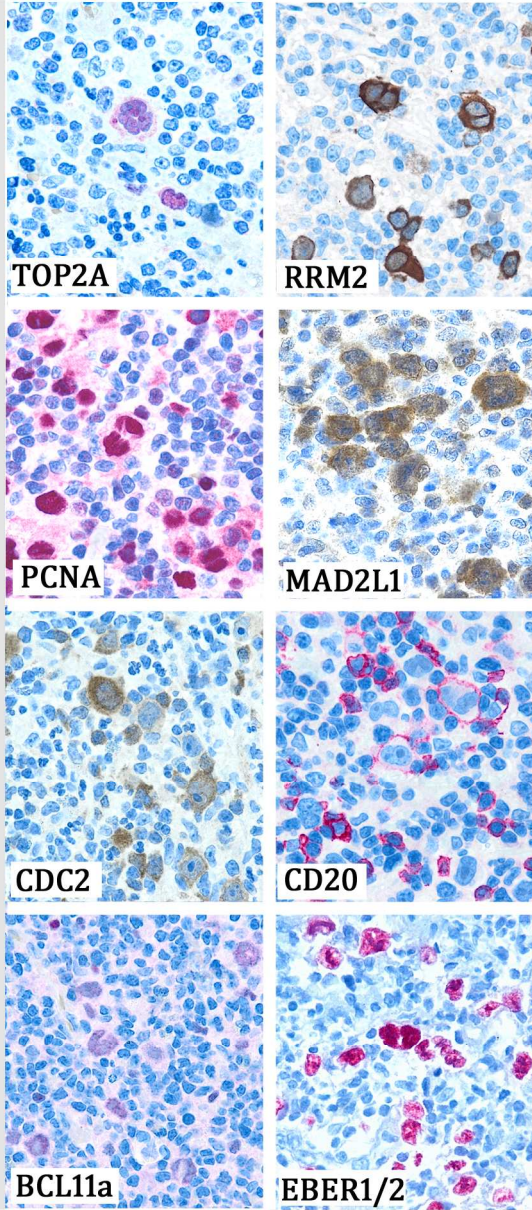


# PET-2/Biologic Markers Study

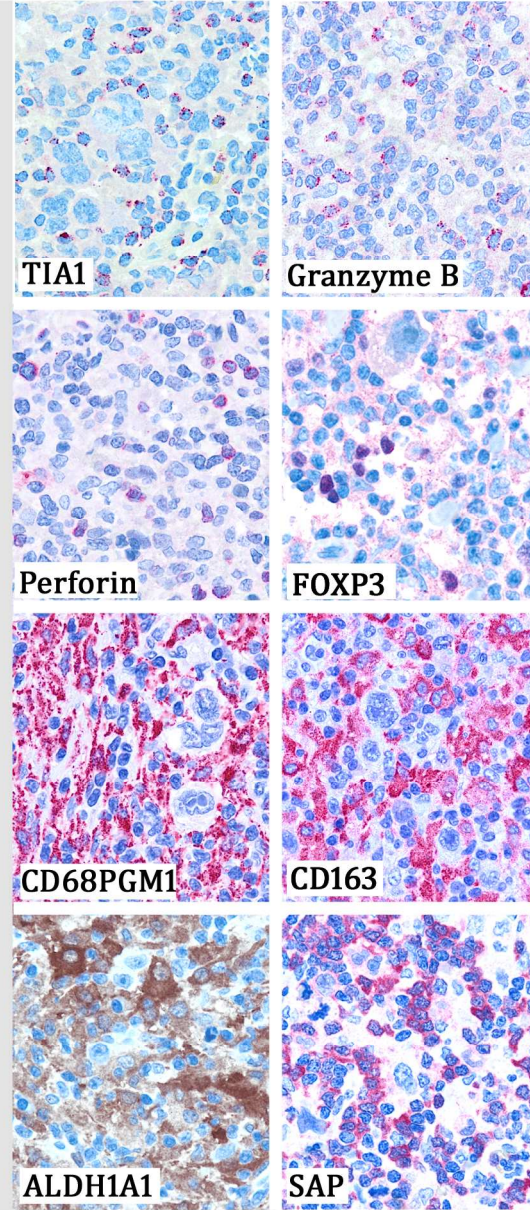
- Biopsy samples from cHL patients at diagnosis enrolled by 11 Italian and 1 Danish medical institutions and Polish Lymphoma Research Group allied centers
- Construction of TMAs to collect cases of interest in the same block and optimization of immunohistochemical procedures
- We thoroughly investigated a wide set of biological markers, representative of diverse key aspects of cHL neoplastic and bystander cell biology : cell cycle regulatory proteins (**TOP2A, RRM2, MAD2L1, CDC2, PCNA**), B-cell ontogeny-related proteins (**BCL11a, CD20**), cell damage and apoptosis markers (**P53, BCL2**), EBV infection status, and on macrophages related markers (**CD68, CD163, ALDH1A1, STAT1**) and T-cell cytotoxicity (**TIA1, Perforin, Granzyme B**), regulation and suppression markers (**FOXP3, PD1, SAP**).
- Evaluation of the prognostic impact of such markers on Hodgkin's lymphoma outcome
- We challenged their prognostic/predictive power versus interim PET
- Construction of a predictive model of lymphoma recurrence



**clone-related markers**



**microenvironment-related markers**



# PET-2/Biologic Markers Study

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- We thoroughly investigated a wide set of biological markers, representative of diverse key aspects of cHL neoplastic and bystander cell biology : cell cycle regulatory proteins (**TOP2A**, **RRM2**, **MAD2L1**, **CDC2**, **PCNA**), B-cell ontogeny-related proteins (**BCL11a**, **CD20**), cell damage and apoptosis markers (**P53**, **BCL2**), EBV infection status, and on macrophages related markers (**CD68**, **CD163**, **ALDH1A1**, **STAT1**) and T-cell cytotoxicity (**TIA1**, **Perforin**, **Granzyme B**), regulation and suppression markers (**FOXP3**, **PD1**, **SAP**).
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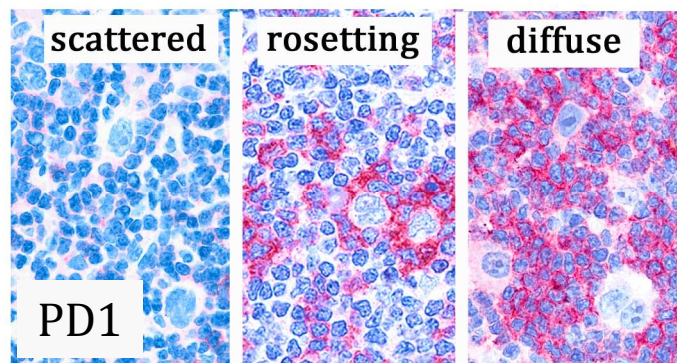
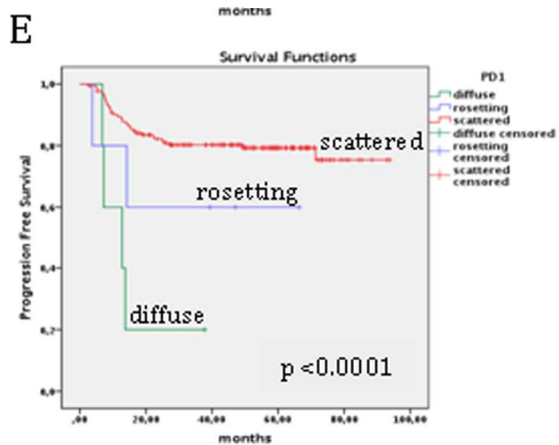
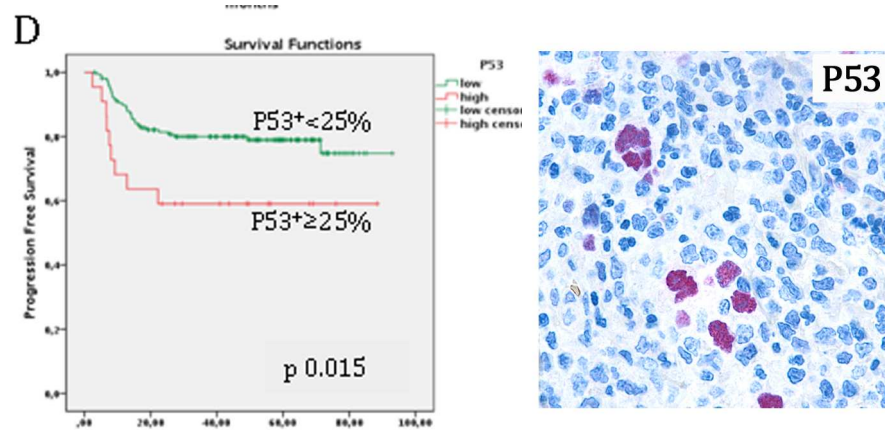
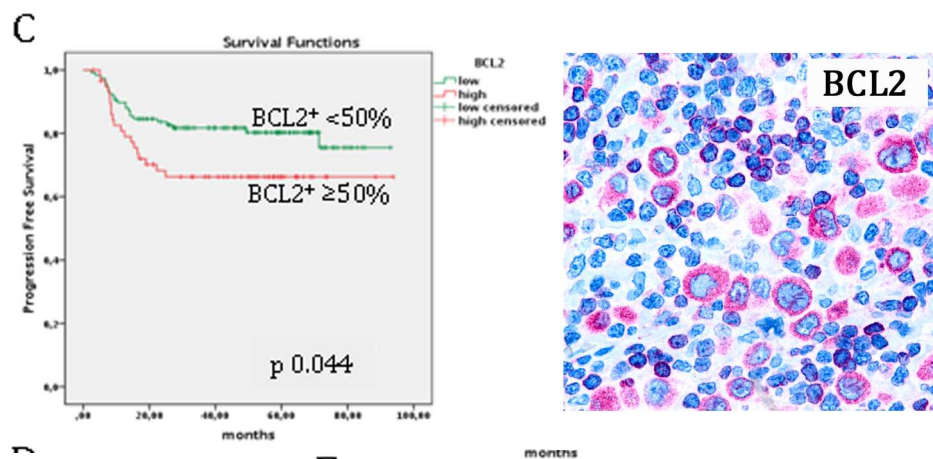
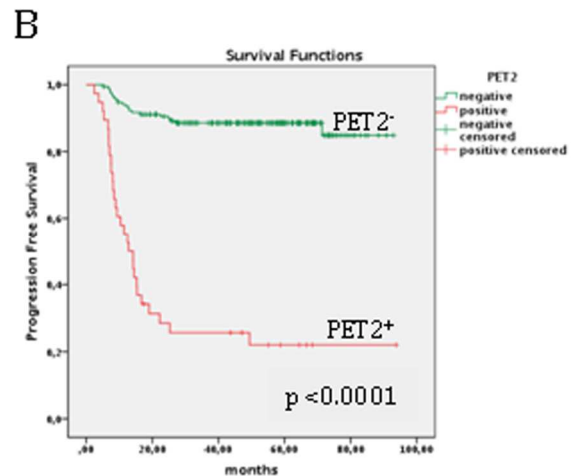
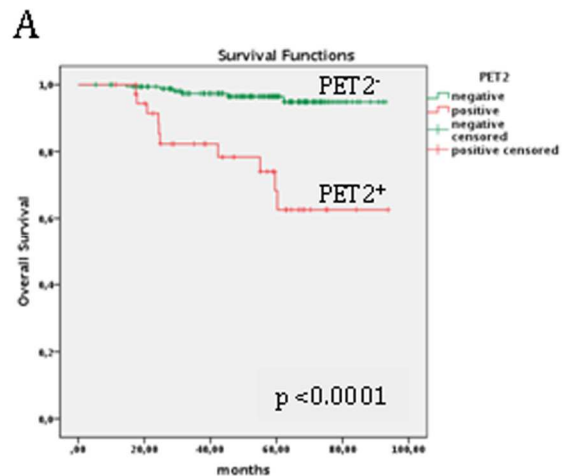


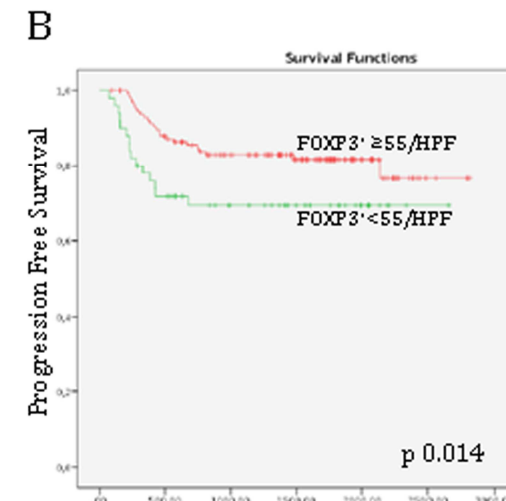
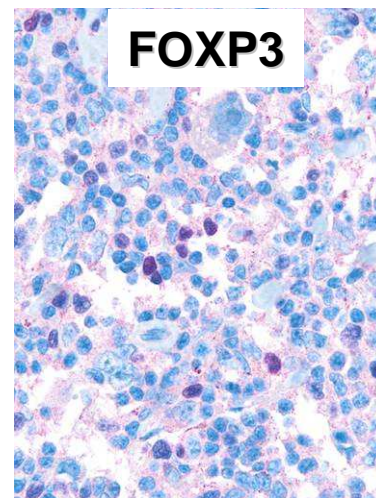
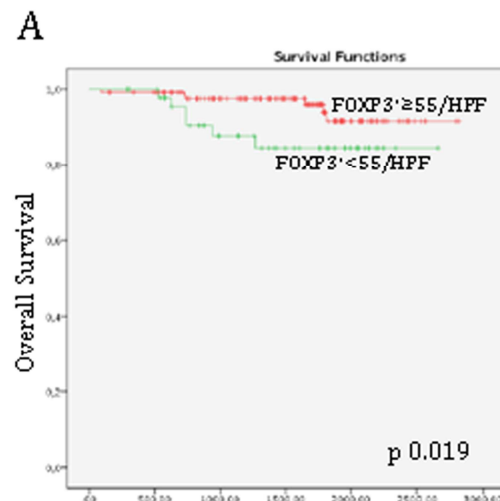
## 310 Patients: 208 pts + 102 pts

|                            | Training set<br>208 pt | Test set<br>102 pt |
|----------------------------|------------------------|--------------------|
| <b>Age</b>                 |                        |                    |
| mean                       | 36                     | 36                 |
| median                     | 32                     | 31                 |
| range                      | 14-80                  | 19-79              |
| <b>Sex</b>                 |                        |                    |
| male                       | 103 (49,5%)            | 52 (51,5%)         |
| female                     | 105 (50.5%)            | 51(48,5%)          |
| <b>Follow-up months</b>    |                        |                    |
| mean                       | 50.26                  | 68.28              |
| median                     | 52.30                  | 68.1               |
| range                      | 3-93                   | 9.9-140.7          |
| <b>Histologic subtype*</b> |                        |                    |
| cHL/nos                    | 9 (9.1%)               | 3 (3%)             |
| NS/nos                     | 9 (4.3%)               | 3 (3%)             |
| NS-cellular phase          | 9 (4.3%)               | -                  |
| NS-1                       | 93 (44.7%)             | 44 (43.1%)         |
| NS-2                       | 37 (17.9%)             | 13 (12.7%)         |
| NS-syncytial variant       | 8 (3.8%)               | 4 (3.9%)           |
| MC                         | 29 (14%)               | 30 (29.4%)         |
| LR                         | 1 (0.5%)               | 5 (4.9%)           |
| LD                         | 3 (1.4%)               | -                  |

|                             | Training set<br>208 pt | Test set<br>102 pt |
|-----------------------------|------------------------|--------------------|
| <b>Ann Arbor stage</b>      |                        |                    |
| I                           | 9 (4.3%)               | 6(5.9%)            |
| IIA                         | 68 (32,7)              | 18 (17,6)          |
| IIB                         | 49 (23,6)              | 15 (14,7)          |
| III                         | 49 (23.6%)             | 26 (25.5%)         |
| IV                          | 33 (15.9%)             | 37 (36.3%)         |
| <b>Symptoms</b>             |                        |                    |
| A                           | 110 (53%)              | 38 (37%)           |
| B                           | 98 (47%)               | 64 (63%)           |
| <b>Bulky disease</b>        |                        |                    |
| Yes                         | 42 (20.1%)             | 42 (41%)           |
| No                          | 166 (79.9%)            | 60 (59%)           |
| <b>First-line treatment</b> |                        |                    |
| ABVD                        | 208 (100%)             | 102 (100%)         |
| RT                          | 52 (25%)               | 43 (42%)           |
| <b>PET after two cycles</b> |                        |                    |
| Negative                    | 170 (81.7%)            | 79 (77.5%)         |
| Positive                    | 38 (18.3%)             | 23 (22.5%)         |
| <b>Clinical outcome</b>     |                        |                    |
| Failure                     | 49 (23.6%)             | 22 (21.6%)         |
| Death of disease            | 16 (7,7%)              | 4 (3,9)            |







**C** Cox's regression model: Overall Survival

| variable | Hazard Ratio of event risk | 95% CI for HR | p     |
|----------|----------------------------|---------------|-------|
| PET-2+   | 0.032                      | (0.04-0.274)  | 0.002 |

**D** Cox's regression model: Progression Free Survival

| variable              | Hazard Ratio of event risk | 95% CI for HR   | p     |
|-----------------------|----------------------------|-----------------|-------|
| Stage II              |                            |                 | 0.001 |
| Stage III             | 2.138                      | (1.756-2.522)   | 0.004 |
| Stage IV              | 3.562                      | (3.235-3.888)   | 0.003 |
| FOXP3 <sup>high</sup> | 0.324                      | (0.147-0.715)   | 0.005 |
| P53 <sup>+</sup>      | 2.624                      | (1.043-6.579)   | 0.040 |
| PET-2+                | 33.333                     | (13.513-83.333) | 0.000 |

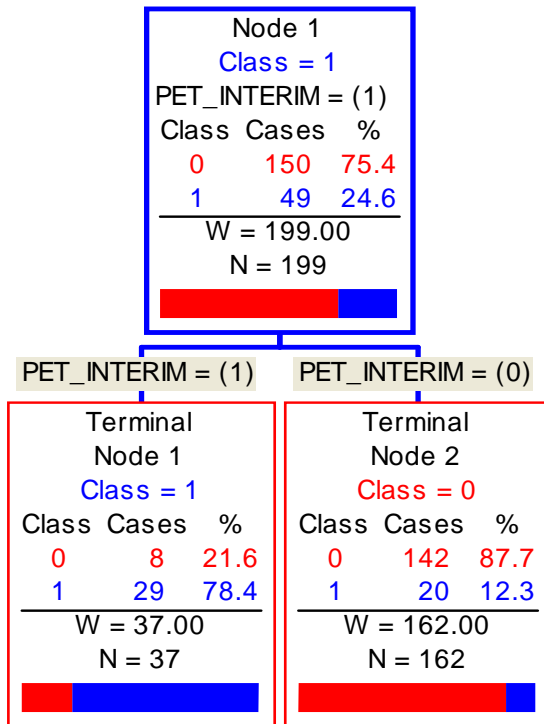
**E** Progression Free Survival after 5 years for different categories

| Variable | Category          | PFS    |
|----------|-------------------|--------|
| Stage    | Stage=2           | 74.12% |
| Stage    | Stage=3           | 59.46% |
| Stage    | Stage=4           | 52.17% |
| FOXP3    | Low: $< 55/HPF$   | 58.83% |
| FOXP3    | High: $> 55/HPF$  | 76.34% |
| P53      | Low: $< 25\%$     | 71.43% |
| P53      | High: $\geq 25\%$ | 50.00% |
| PET2     | Negative          | 83.33% |
| PET2     | Positive          | 14.71% |

# Classification and Regression Tree (CART) analysis

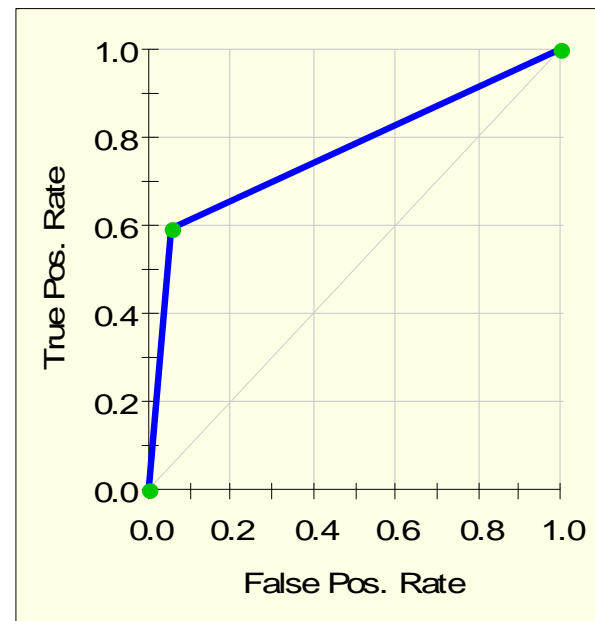
## Misclassification for Learn and **Test** Data

### PET-2 alone based model

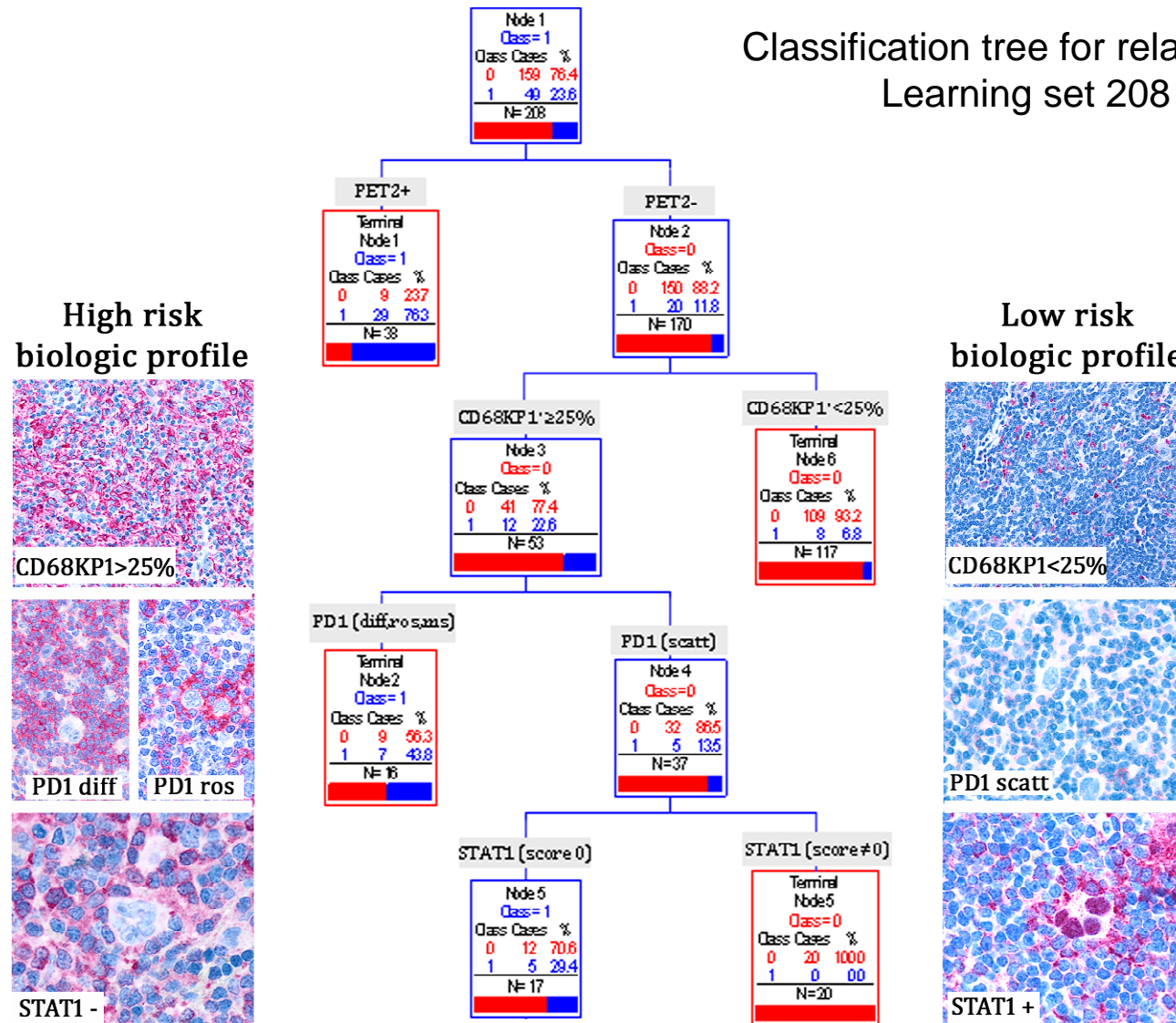


| Class | N Cases | N Mis-Classed | Pct Error |
|-------|---------|---------------|-----------|
| 0     | 150     | 8             | 5.33      |
| 1     | 49      | 20            | 40.82     |
| Tot   | 199     | 28            | 14.07     |

ROC Integral: 0.770



## Classification tree for relapse/progression Learning set 208 patients



### low-risk class:

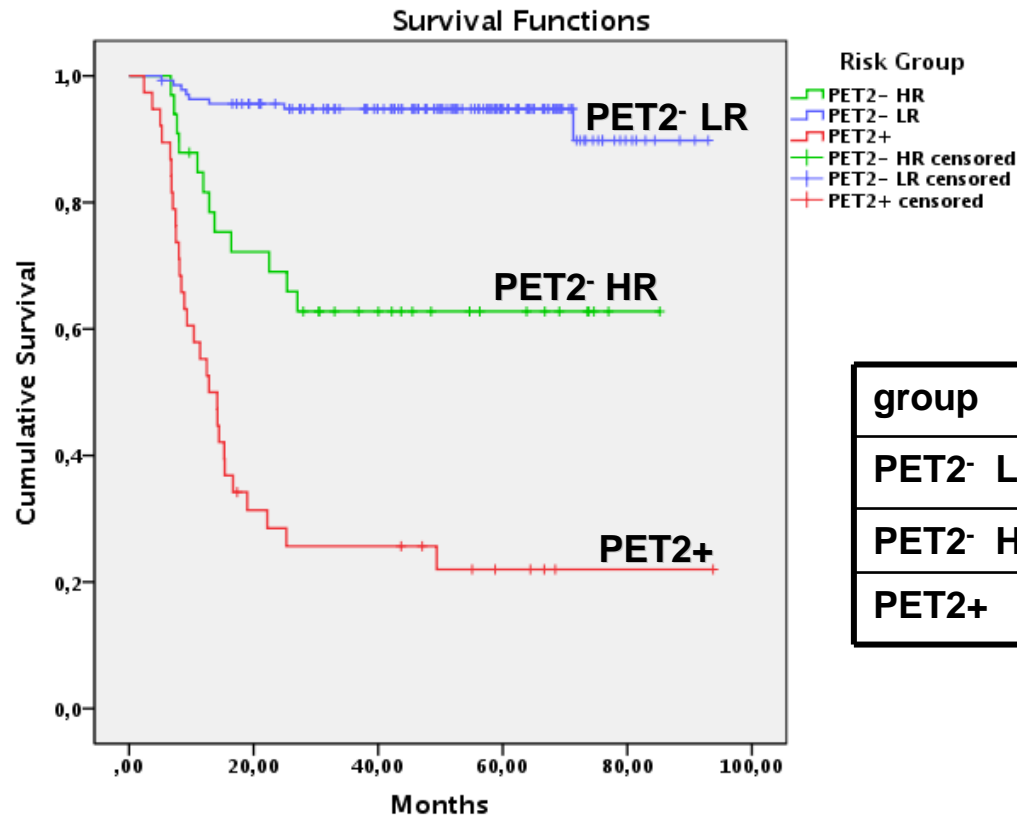
- patients that, in addition to a negative PET-interim, had low values of CD68KP1 percentage  $< 25\%$  ;
- patients that, in addition to a negative PET-interim, and despite CD68KP1  $\geq 25\%$ , had jointly scattered PD1 pattern and positive STAT1 in RCS;

### high-risk class:

- patients that, despite a negative PET-interim, were characterised by CD68KP1  $\geq 25\%$  and diffuse or rosetting PD1 pattern;
- subjects that, despite a negative PET-interim, had jointly CD68KP1  $\geq 25\%$ , scattered PD1 pattern and were negative for STAT1.

# misclassification rates

| Class | Pet-2 based classification scheme in learning set: 208 pts |               |                   | Pet-2 + biologic markers classification scheme         |               |                   |                        |               |                   |
|-------|--|---------------|-------------------|--|---------------|-------------------|------------------------|---------------|-------------------|
|       |  |               |                   | Learning set: 208 patients<br>10-fold cross validation |               |                   | Test set: 102 patients |               |                   |
|       | number of cases  | n of misclass | misclass rate (%) | number of cases  | n of misclass | misclass rate (%) | number of cases        | n of misclass | misclass rate (%) |
| 0     | 159  | 9             | 5.66%             | 159  | 26            | 16.35%            | 80                     | 10            | 12.50%            |
| 1     | 49   | 20            | 40.82%            | 49   | 11            | 22.45%            | 22                     | 3             | 13.63%            |
| total | 208  | 29            | 13.94%            | 208  | 37            | 17.79%            | 102                    | 13            | 12.74%            |



| group    | 5 yrs PFS |
|----------|-----------|
| PET2- LR | 94,7%     |
| PET2- HR | 63,6%     |
| PET2+    | 23,7%     |

# Conclusions

- none of the biologic factors evaluated could perform better than PET-2 in predicting outcome.
- a clinical-pathological algorithm based on the combination of PET-2 with some of the investigated biological markers, allowed the identification of poor prognosis patients that were misclassified by PET-2 alone.

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